

“Best Practices for Bioassay Testing of Food and other Complex Mixtures”

Summary of Amsterdam
Workshop

Dr. Thomas Gude



AOAC Europe Section

- **Our Mission**

- AOAC Europe brings together government, industry, and academia to establish standard methods of analysis that ensure the safety and integrity of foods and other products that impact public health around the world

- **What you do not know**

- Start in 1983
- As a member of AOAC International you are automatically a member of AOAC Europe section – when living/working in Europe

- **What you should know**

- We are organizing regularly meetings together with other European Societies
- We are organizing workshops:
 - Nov 2023: Bioassay
 - Q3 2024: Non-Target Methods

- **Please check: <https://aoaceurope.com/>**

The Amsterdam Meeting

What we like to achieve:

- In Amsterdam:

- **Awareness on the topic**
- **Commitment for fruitful discussions**
- **An initial working plan, which topics needs to be addressed**

- Today:

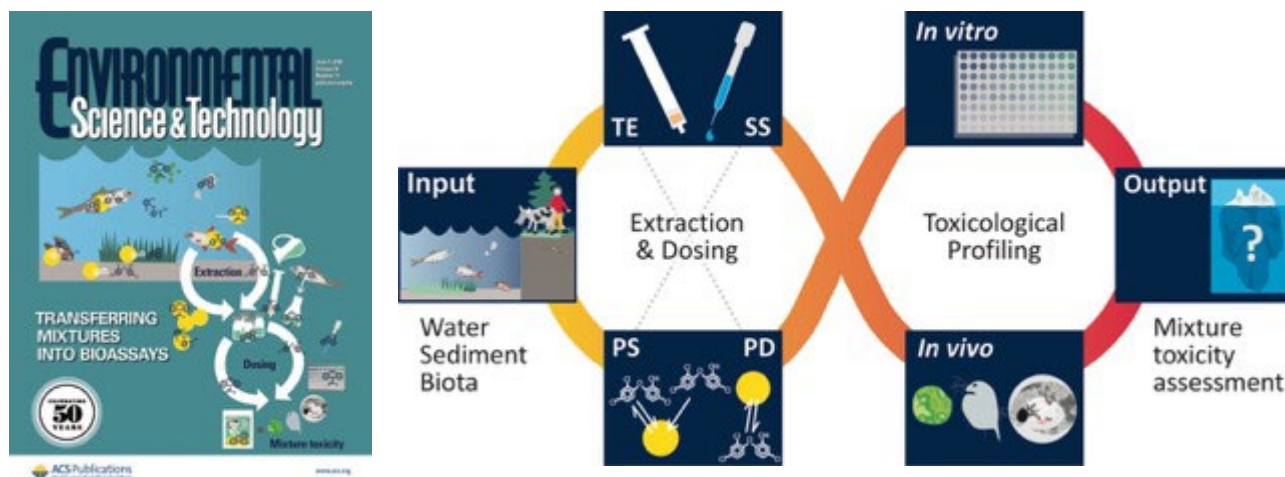
- **Working group(s) covering sub-questions**
- **Harmonized Guideline/Method on “Best Practices for Bioassay Testing of Food and other Complex Mixtures”**

Today's Program



- 05:00 PM General Introduction by AOAC Europe and Workshop details**
- 05:10 PM Summary of Talks given in Amsterdam**
- 1: Prof. Beate Escher – Presenter Georg Braun**
Title presentation: QA/QC during applications of *in vitro* assay for food monitoring
 - 2: Prof. Bennard van Ravenzwaay – Presenter Maricel Marin-Kuan**
Title presentation: The use of metabolomics as a New Approach Methodology (NAM) in the context of the transition from *in vivo* to *in vitro* methods
 - 3: Dr. Ir. Toine Bovee – Presenter Maricel Marin-Kuan**
Title presentation: Development, validation and application of bioassays: their added value.
 - 4: Dr. Peter A. Behnisch – Presenter Peter Behnisch**
Title presentation: Plastic and plastic additives testing by effect-based bioanalysis for endocrine disrupting chemicals – a CRO perspectives
- 05:30 PM Summary of Break Out Groups Amsterdam meeting– Presenter Thomas Gude**
- 05:45 PM AOAC Bioassays Working group proposal: Guidelines on Best Practices for Bioassay Testing of Food and other Complex Mixtures (Presenter Maricel Marin-Kuan)**
- 06:15 PM AOAC International and Bioassays working group meeting (Presenter Kate Mastovska)**
- 06:30 PM End**

QA/QC during applications of *in vitro* assay for food monitoring



Beate I. Escher and Georg Braun

Cell Toxicology, Helmholtz Centre for Environmental Research – UFZ, Germany
 Eberhard Karls University Tübingen, Germany

QA/QC for application for bioassays for food monitoring

Not all bioassays that work for single chemicals are **amendable** to testing of **extracts**

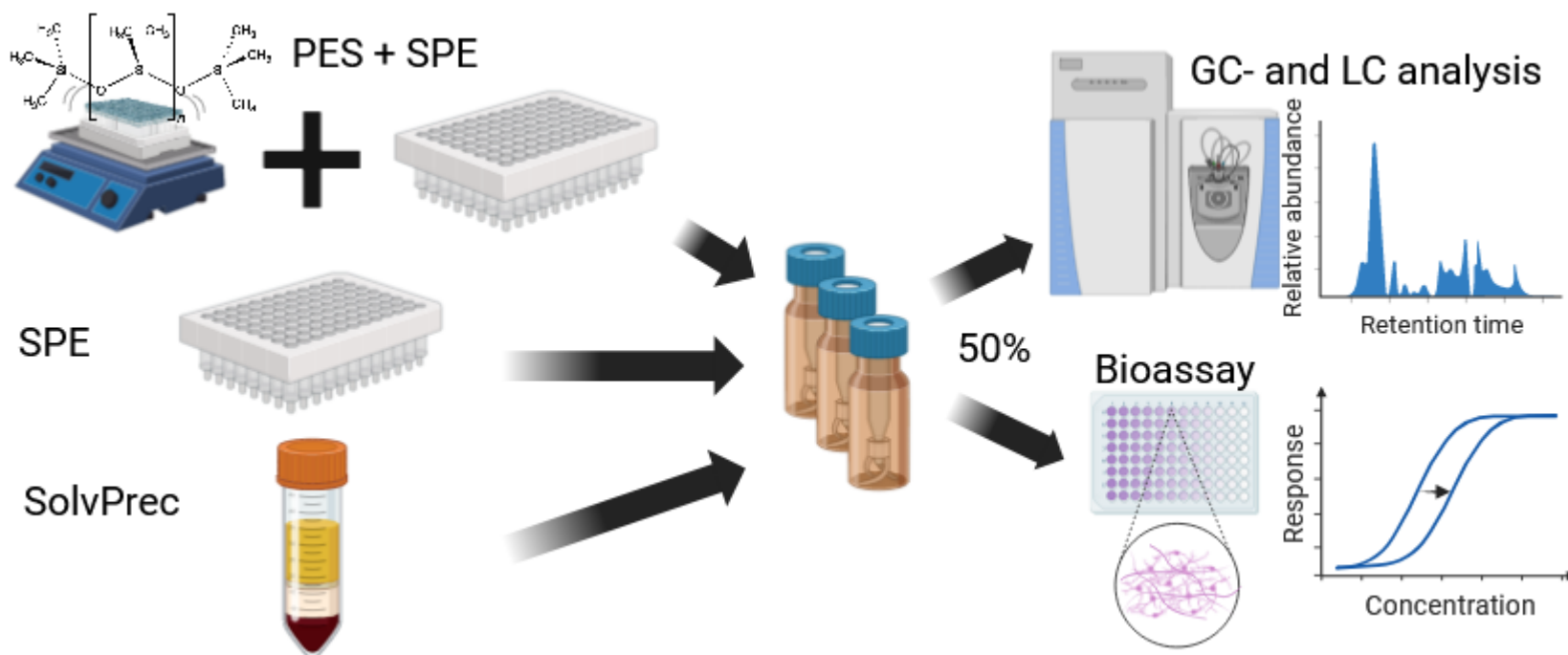
- **Coextracted matrix and endogenous compounds** might interfere with the bioassay
 - **Controls:** Processing blanks, effect recovery of spiked samples
- The more difficult the sample the better must be the bioassay quality
- **Extraction methods crucial** (poor sample extract – poor bioassay result)

Extraction must be **unbiased and independent on the physicochemical properties** of the chemical mixtures to assure that the dosed mixture is representative of the sample

- Extraction method should show commonalities for liquid and condensed phases
- Coextracted matrix (= lipids, proteins) must be minimised
- Internal standards for extraction recovery not possible: independent recovery experiments necessary

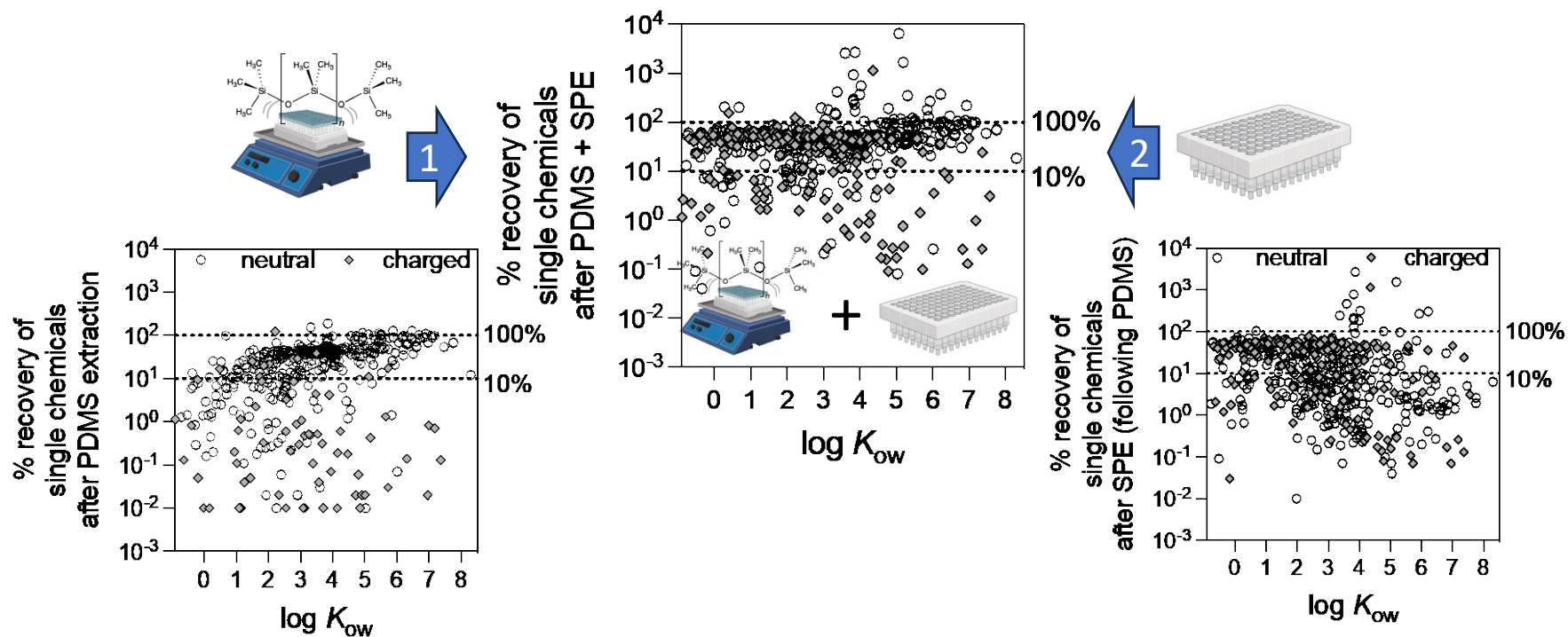
Mixtures of spiked chemicals extracted from blood plasma

- Blood, plasma, serum, food matrices like cow milk or plant-based milk are protein-rich with water content
- Typical used solid-phase extraction neglects the highly bioactive hydrophobic chemicals
- Comparison of recovery of chemicals (n=382) between PDMS and PDMS followed by SPE: good recovery of 2-step method



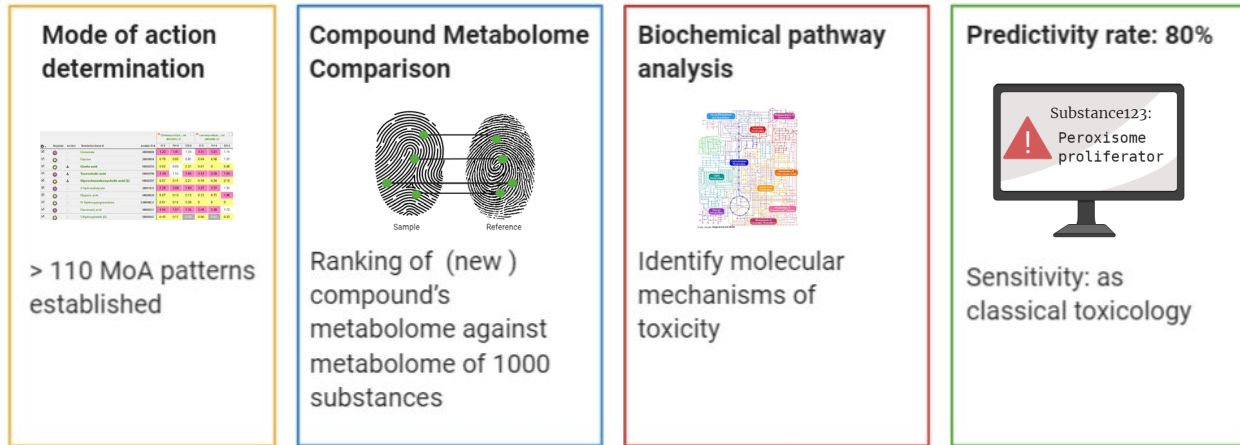
Recovery of mixtures of chemicals extracted from blood plasma

Two-step method achieves best recovery for all chemicals with exception of cations



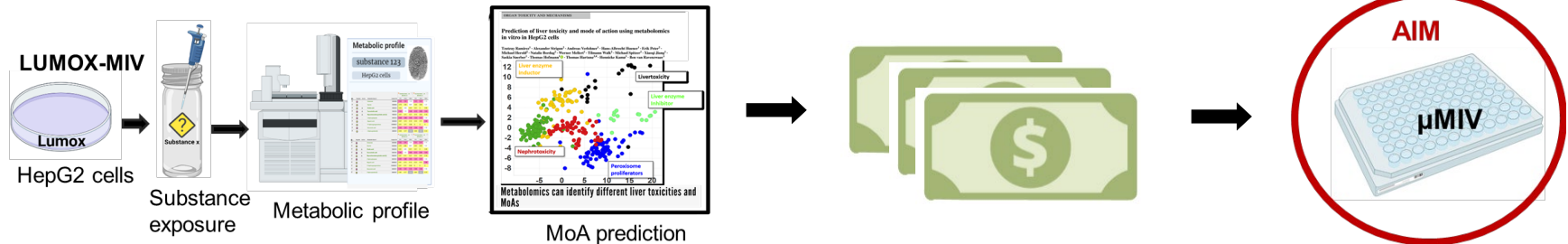
Metabolomics from in vivo to in vitro

Prof. Dr. Bennard van Ravenzwaay



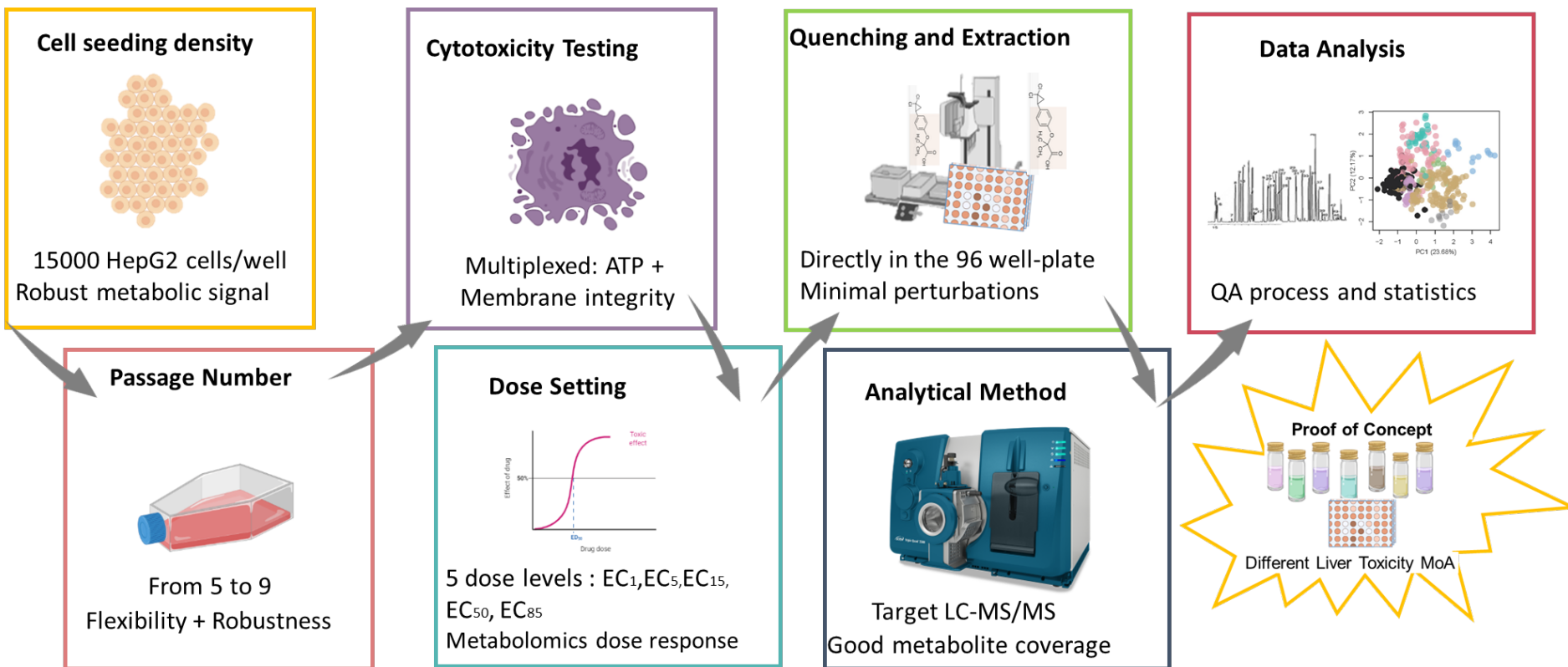
MetaMap[®]Tox:
 an in vivo success story
 van Ravenzwaay et al Tox Letters, 172, 2007

Metabolomics in vitro works.... Ramirez, T. et al. 2018, Archives of Toxicology, 92 *but*



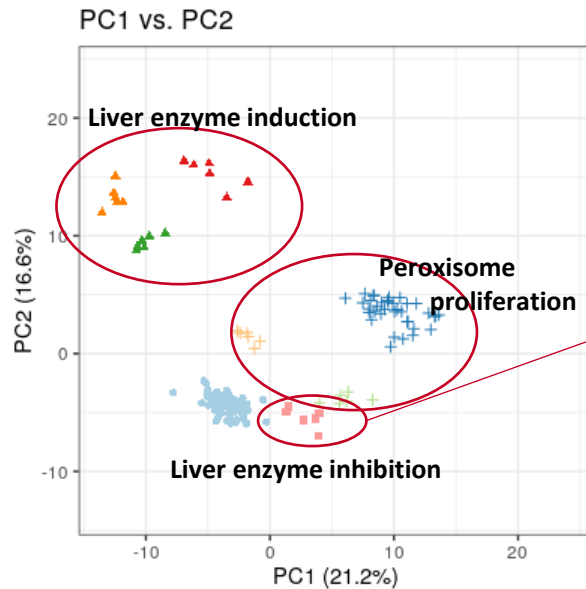
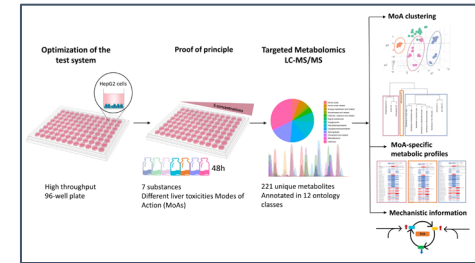
Miniaturization of Metabolomics in vitro

Prof. Dr. Bennard van Ravenzwaay



Proof of principle of the μ MIV platform – μ MIV distinguishes different Mode of Actions

μ MIV identifies different toxicological Mode of Actions

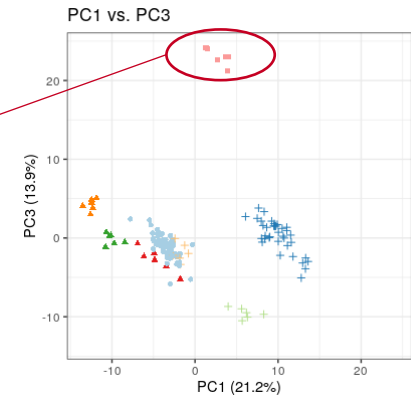


Treatment

- Control
- Bezafibrate
- Acicfluorfen C3
- Aroclor 1254 C2
- Ketoconazole C3
- Pendimethalin C3
- Wy-14643 C3
- β -Naphthoflavone C3

MoA cluster

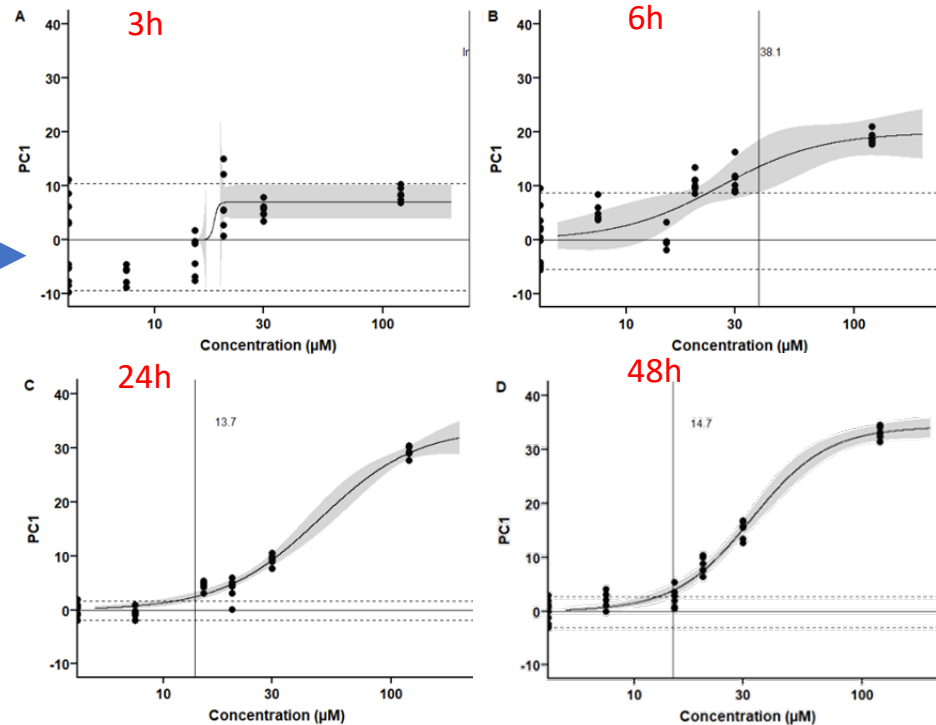
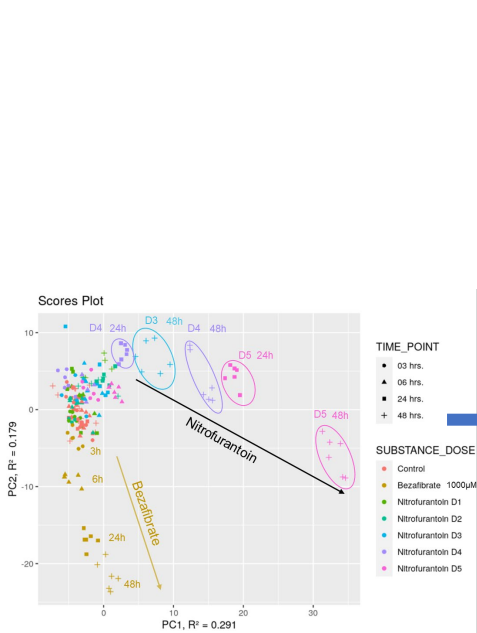
- Control
- ▲ Liver enzyme induction
- Liver enzyme inhibition
- +



MoA = Mode of action
 μ MIV = miniaturization of metabolomics in vitro

Ramirez-Hincapie, S. et al. 2023, Cell biology and toxicology, open access

Point of departure (PoD) derivation from μ MIV data at different time points (3h, 6h, 24h and 48h)



The PoD at earlier time points is higher (less „toxic“)

At 24h and 48h the PoD approaches a similar value.

Exposure time plays a role for PoD derivation.

Ramirez-Hincapie et al. 2023, Archives of Toxicology, 97, open access

PoD = Point of departure
 μ MIV = miniaturization of metabolomics *in vitro*

Reduction, Refinement, Replacement

Metabolomics *in vivo* and *in vitro*: an overview of applications

- | | |
|--|-------------------------|
| 1) Screening: early detection of „bad“ compounds | Tox. Letters, 172, 2007 |
| 2) Identification of molecular mechanisms | Tox. Letters, 225, 2014 |
| 3) Chemical grouping and read across
Reg. Tox. Parm. 81, 2016 | Mut. Res. 746, 2012, |
| 4) Smart Studies: include omics data in regulatory testing | Arch. Tox. 96, 2022 |
| 5) <i>In vitro</i> metabolomics (MIV) | Arch. Tox., 92, 2018 |
| 6) μ MIV: hazard and MoA identification | Cell biol. Tox. 2023 |
| 7) μ MIV: dose and time response, PoD | Arch. Tox., 97, 2023 |

Take-home messages:

- ✓ Growing demonstration of feasibility for *in vitro* application
- ✓ Multiple mode of action can be identified with one test
- ✓ This technology allows for determination of point of departure

Development, validation and application of bioassays: their added value

Toine Bovee
November 3, 2023
Amsterdam



Bioassays for food control, there are many:

- 1) **Cell based effect bioassays**, e.g. for the detection of antibiotics, dioxins and PCBs, steroid hormones, marine toxins, lectins, and mutagenic compounds
 - 2) **Enzyme based “effect bioassays”**, e.g. for the detection of microcystins, statins, NSAIDs, pesticides and Viagra-like compounds
 - 3) **Other bio-based methods**, e.g. TTR, TBG, and β 2-adrenoceptor competitive binding assays, or based on a whole organism, like the DapTox test with Daphnia
- Once developed, and proven to work with academic standards, these **assays need to be made applicable for a certain matrix**. I.e., methods need to be developed for e.g. animal feed, supplements, water, milk, egg, meat, urine, mussels, fish, hair and feathers.
 - After method development for a certain matrix or several matrices, these **methods need to be validated and accredited before they can be used in routine monitoring for food control purposes with legal strength**.

Bioassays in food control, their added value

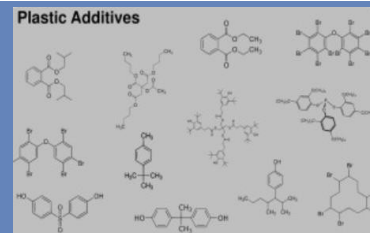
- Mass spectrometric analysis is often targeted and of great strength for enforcement. However, even untargeted MS methods are most often not able to detect unknown bioactives. Bioassays, and especially the cell-based effect assays, detect both the known contaminants/toxins and the yet unknown toxins (unknown bio-actives). The latter is an added value for food safety control. There are many examples where bioassay analysis has shown this added value compared to MS analysis only. E.g.:
 - detection of estradiol in animal feed (with the yeast estrogen bioassay)
 - detection of DES in human supplement (with the yeast estrogen bioassay)
 - detection of 1-testosterone, 4-androstenediol and 5-androstanediol in human supplements (with the yeast androgen bioassay)
 - detection of THG (tetrahydrogestrinone) in human urine (with the yeast androgen bioassay)
 - detection of TBDF (tetrabromodibenzofuran) in animal feed, egg, and chicken meat (with the DR-CALUX bioassay)

Bioassays in food control, their added value

- detection of Cl-PAH in chlorinated paraffins (with the DR-CALUX bioassay)
- detection of dexamethasone in human supplement (with the yeast (gluco)corticoid bioassay and GR-CALUX bioassay)
- detection of ciguatoxin in fish (with the neuro-2a bioassay)
- detection of sildenafil analogues in human supplements (with the PDE-5 enzyme inhibition assay)
- detection of isopropyloctopamine in human supplement (with the competitive β -adrenoceptor binding assay)

**Thank you
for your
attention!**

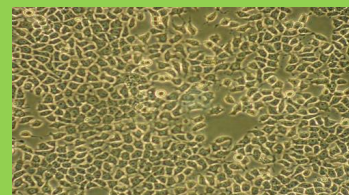
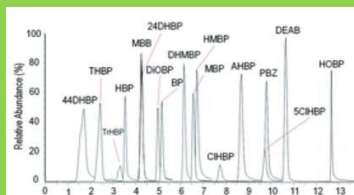




Plastic, water and chemicals testing by non-animal methods (NAMs) – a CRO's perspective

Peter A. Behnisch and Harrie Besselink

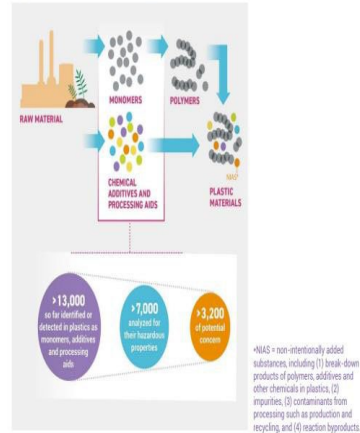
BioDetection Systems, Amsterdam, The Netherlands





Unknown amount of unknown chemicals with unknown in vitro toxicity profiling – Urgent steps needed...(UN, June 2023)

Figure 3. A simplified illustration of the production of plastics from raw materials, and an overview of chemicals that may have been used in plastic production or have been detected in plastics.



Chemicals of concern in plastics are found across various sectors and products value chains

SOLUTIONS to move FORWARD

Update regulatory testing guidelines, for instance by including rapid and cost-effective approaches such as bioassays and computational tools. This serves as a prerequisite for research to fill existing knowledge gaps, e.g., regarding the mixture toxicity of leachates from plastics or the toxicity and bioaccumulation potential of hydrophobic chemicals.

One option to assess the overall toxicity of chemicals released from plastic products is to test the overall extract/leachate using in vitro bioassays, which are rapid and cost-effective (Groh and Muncke 2017), and use other screening technologies that measure relevant toxicological effects such as cytotoxicity, genotoxicity and endocrine effects (Table 4) (Koster et al. 2016). The AOP approach (see section 4.1.2) can



Table 4. A selection of in vitro bioassays that have been used in NIAS research (Koster et al. 2016).

Potential endocrine activity	Cytotoxicity	Genotoxicity/potential carcinogenicity
Oestrogen receptor (ER) redistribution Androgen receptor (AR) redistribution	Cell Organelle Health (COH); end points: DNA content, cytochrome C, mitochondrial membrane potential, RNA synthesis kinetic inhibition.	Indicator assays for genotoxicity (PARP, GADD45...), Comet-FPG assay
Transcriptional activation assay	Cell Proliferation and Cell Death (CPD); end points: apoptosis – caspase3, p53; DNA content, DNA proliferation – BrdU	Mutagenicity test (Ames test, mammalian cell gene mutation tests, micronucleus (MN) test)
Oestrogen receptor (ER) (anti) Androgen receptor (AR) Glucocorticoid receptor (GR) Progesterone receptor (PR) Thyroid receptor (TR) Peroxisome Proliferator Activated Receptor (PPAR γ)		Potential carcinogenicity Cell Transformation Assay (detection of both geno- and non-genotoxic carcinogens)
H295R Steroidogenesis assay (changes in hormone production)		



Known vs Unknown Chemicals – Urgent steps needed...

Muncke et al. *Environmental Health* (2020) 19:25
<https://doi.org/10.1186/s12940-020-0572-5>

Environmental Health

COMMENTARY

Open Access

Impacts of food contact chemicals on human health: a consensus statement

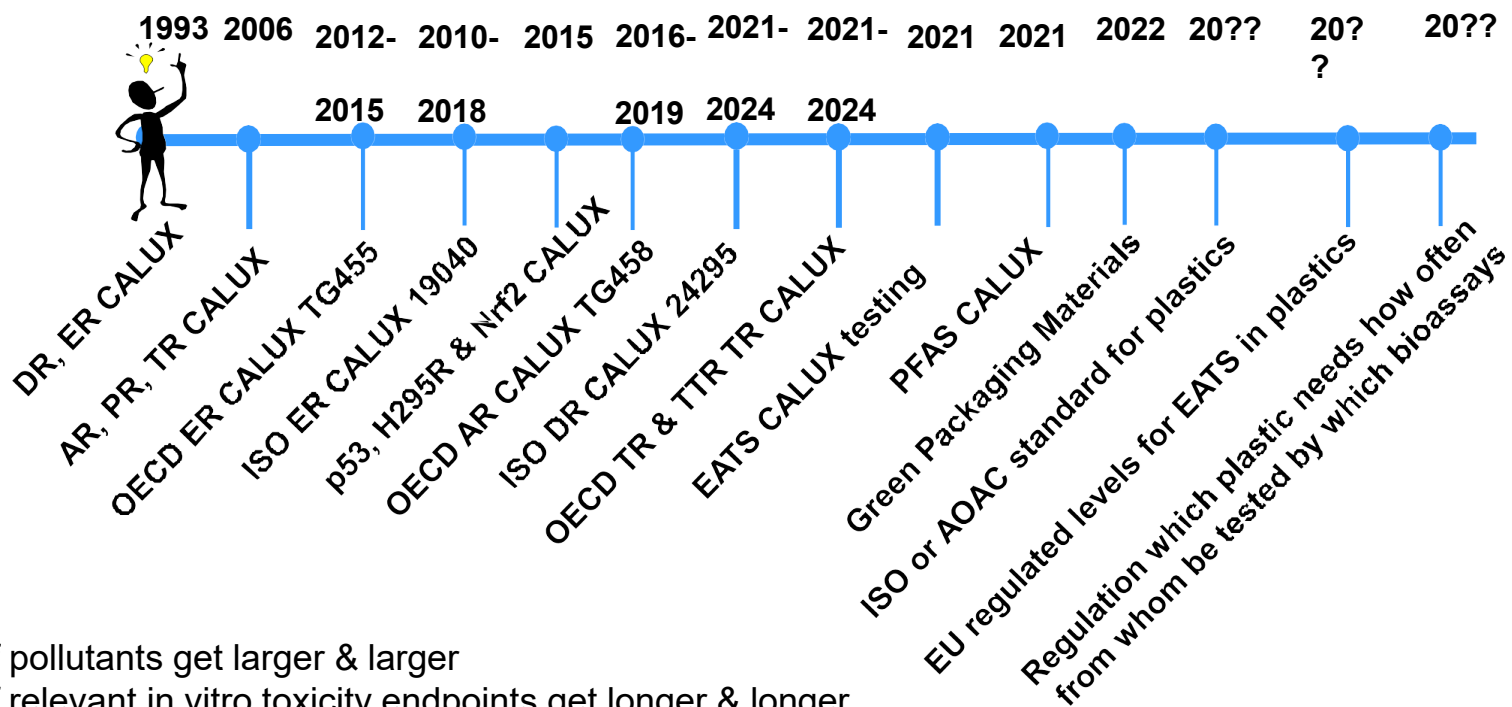


Jane Muncke^{1*}, Anna-Maria Andersson², Thomas Backhaus³, Justin M. Boucher⁴, Bethanie Carney Almroth³, Arturo Castillo Castillo⁵, Jonathan Chevrier⁶, Barbara A. Demeneix⁷, Jorge A. Emmanuel⁸, Jean-Baptiste Fini⁷, David Gee⁹, Birgit Geueke¹, Ksenia Groh¹, Jerrold J. Heindel¹⁰, Jane Houlihan¹¹, Christopher D. Kassotis¹², Carol F. Kwiatkowski¹³, Lisa Y. Lefferts¹⁴, Maricel V. Maffini¹⁵, Olwenn V. Martin¹⁶, John Peterson Myers^{17,18}, Angel Nadal¹⁹, Cristina Nerin²⁰, Katherine E. Pelch¹³, Seth Rojello Fernández²¹, Robert M. Sargis²², Ana M. Soto²³, Leonardo Trasande²⁴, Laura N. Vandenberg²⁵, Martin Wagner²⁶, Changqing Wu²⁷, R. Thomas Zoeller²⁸ and Martin Scheringer^{4,29}

- **Toxicity** and exposure information is available **only for few of the IAS/NIAS**
- Risk assessment **of unknown chemicals is not possible** under the current regulatory approach
- **Modernize tiered approach for screening and prioritization**
- Addressing **mixture toxicity**
- Modernizing risk assessment by including **endocrine disruption**



Key milestones of bioassays (NAMs) for developing guidance docs (e.g., for plastic)



- list of pollutants get larger & larger
- list of relevant in vitro toxicity endpoints get longer & longer
- list of the be tested materials get larger & larger

➡ standardization process for EDCs in plastic materials needs more attention!



Endocrine Disrupting Chemicals by EATS-NAMs testing

Typical case study

Analysis of compounds suspect of endocrine disruption (EDC) on a dedicated **EATS**-panel:

- (anti-) **E**strogenicity
- (anti-) **A**ndrogenicity
- (anti-) **T**hyroidogenicity
- T**TR binding
- T**hyroid peroxidase (hTPO) inhibition
- S**teroidogenesis (H295R)

Read-across and new approach methodologies applied in a 10-step framework for cosmetics safety assessment – A case study with parabens

Gladys Ouedraogo^a, Camilla Alexander-White^b, Dagmar Bury^c, Harvey J. Clewell III^d, Mark Cronin^e, Tom Cull^f, Matthew Dent^f, Bertrand Desprez^g, Ann Detroyer^a, Corie Ellison^h, Stefania Giammancoⁱ, Eric Hack^j, Nicola J. Hewitt^k, Gerry Kenna^l, Martina Klaric^g, Reinhard Kreiling^m, Cathy Lesterⁿ, Catherine Mahonyⁿ, Enrico Mombelli^o, Jorge Naciff^h, John O'Brienⁱ, Andreas Schepky^p, Sarah Tozerⁿ, Bart van der Burg^q, Barbara van Vugt-Lussenburg^q, Sharon Stuard^h, Cosmetics Europe^{g, r}

^q BDS, Science Park 406, 1098XH, Amsterdam, the Netherlands



G. Ouedraogo et al.

Regulatory Toxicology and Pharmacology 132 (2022) 105161

Table 11

Summary of EATS testing results. PC10 (for agonistic tests)/PC20 (for antagonistic tests) values are shown in -Log M; the color indicates the potency (yellow < orange < red).

Step 7b. Biokinetic refinement

End point	MP		EP		PP		BP		pHBA	
	-S9	+S9	-S9	+S9	-S9	+S9	-S9	+S9	-S9	+S9
Cytotoxicity	>	>	>	>	-3.5	>	-4.0	-3.0	>	>
(Anti-) estrogenic and (anti-) androgenic assays										
ER α CALUX	-4.5	-4.2	-5.5	-4.8	-6.0	-5.1	-6.0	-5.0	>	>
anti-ER α CALUX	>	>	>	>	>	>	>	>	>	>
AR CALUX	>	>	>	>	>	>	>	>	>	>
anti-AR CALUX	-4.9	-4.7	-4.7	-4.4	-4.5	-4.2	-4.9	-4.3	>	>
Thyroidogenic assays										
TR β CALUX	>	>	>	>	>	>	>	>	>	>
anti-TR β CALUX	-3.0	>	>	>	>	>	>	>	>	>
TTR	-2.7	nd	-4.8	nd	-4.5	nd	-4.8	nd	>	nd
hTPO	-2.0	nd	>	nd	>	nd	>	nd	-3.0	nd
Steroidogenesis										
H295R-E2	-5.0	nd	-5.0	nd	-5.0	nd	-5.0	nd	-3.0	nd
H295R-T	>	nd	-4.0	nd	-4.0	nd	>	nd	>	nd



Take home messages

- Since more than 30 years it is known that many plastic additives have hormone- like activities and can be easily picked-up by in vitro bioassays.
- Development of international standards for such in vitro bioassays for endocrine disrupting chemicals are very slowly and have now been finalised for compounds testing (OECD TG 455, 456, 458), for some environmental matrices (ER and DR for water), but not for any materials (such as packaging materials)
- Many published studies show that a panel of bioassays is a safe approach for a greener & sustainable future of plastics & other packaging materials.
- Our studies applied on plastics, water and food matrices shows that they can be easily adopted to many kinds of in vitro toxicity endpoints.
- Now it is urgently needed to step forward to bridge the single chemical compounds testing with the effect-based in vitro biological analysis steps of such complex mixtures of all kinds of known and unknown chemical & toxicological properties.



Summary of Break Out Groups

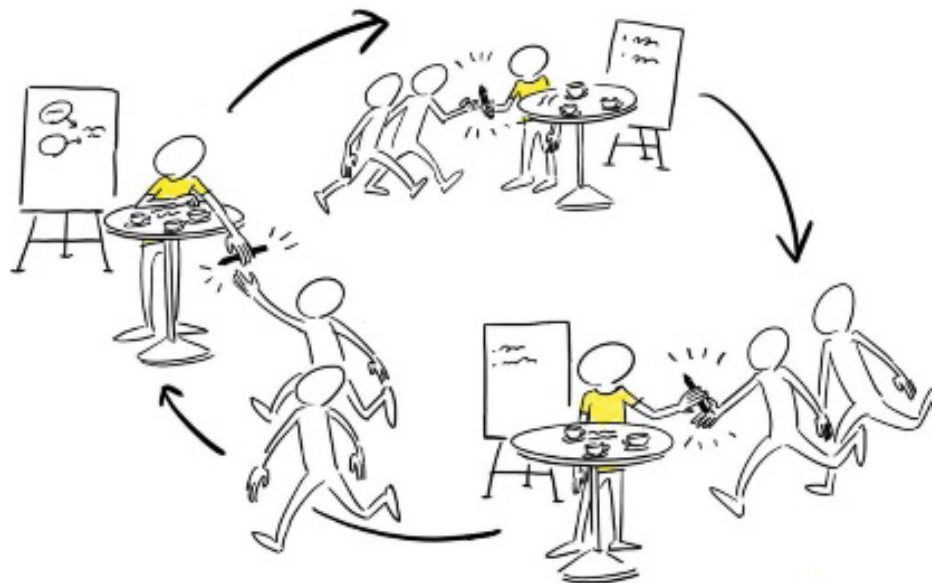
All Participants could join in the “world coffee format” the following 4 stations:

Group 1: OECD Guidance Document on Good In Vitro Method Practices (GIVIMP) use

Group 2: Current extraction practices

Group 3: Critical factors for accurate testing

Group 4: Key topics for future WGs



Group 1: OECD Guidance Document on Good In Vitro Method Practices (GIVIMP) use



Question: Do you apply the OECD Guidance Document on Good In Vitro Method Practices (GIVIMP) and do you consider those as appropriate for complex mixtures? Why are or not appropriate? Other guidelines available to recommend?

Feedback:

- Documentation, standardization and guidelines are important
- Use of control samples
- Validation is slow and costly, but important
- Before the official validation, there should be a platform, where knowledge sharing is facilitated
- Resources (financial, intellectual)
- Single compounds versus mixtures
- More complex
- Chemical analysis and extraction documentation are key

Group 2: Current extraction practices



Question: What consideration are you taking when extracting mixtures from water, food, food contact materials and other items to test with bioassays? (What is best practice and limitation of current approach?)

Feedback:

- Design of controls and blanks
- Consider the behavior of mixtures in your controls (mix instead of single compounds)
- Consider physicochemical properties
- Consider solvents
- Apply different readouts with same mode of action (impact of labels)
- Co-extracts and matrix effects (high recovery vs. interfering matrix)
- Is it beneficial to combine multiple extraction methods to cover a broad range of chemicals?
- Is it beneficial/necessary to explain observed effects?
- Is your extraction compatible in both instrumental analysis and bioanalysis?

Group 3: Critical factors for accurate testing

Question: What are the most critical factors and source of errors for consideration to do accurate testing of complex undefined mixtures and extracts from water, food and related items using bioassays?

Feedback:

1) Sample :

- Type of samples
- Example food simulants for packaging (7 simulants possible)
- Matrix interference generated by the sample?
- Readout interference by sample (fluorescence or protein binding)
- Matrix calibration for bioassays
- Quality controls (spiking test as QA)

2) Extraction:

- Method of extraction (ex. Rotavapor optimization)
- Solubility issues
- Solvents used
- DMSO vs other solvents for cell exposure
- Amount sample to extract
- Concentration issues-volatiles: false-negatives
- Ready to use solvents-long term / harmonization

3) Bioassays:

- Qualified in vitro methods
- Star screening and follow with confirmations.
- Use of DMSO for cell culture exposure?
- Dosing
- Cytotoxicity-LOQ
- Reproducibility
- Concentration LOD
- Matrix effect-particles
- Data interpretation may be bias-interferences/qualification?

4) Specific and non-specific compounds

5) Dominant effect in mixture-check cells viability vs cytotox markers

6) Validation new bioassays to assess FCM

Group 4: Key topics for future WGs

Question: There is a need for development of harmonized best practices (framework) when using bioassays for undefined mixtures.

- a) to your opinion, which are the topics and key procedures that should be covered?
- b) do you think those topics could be addressed through AOAC working groups?

Feedback:

- Potential focus: Food contact materials
- Extraction procedures – solvents (compound extractability vs. biocompatibility with the given bioassay), pre-concentration/dilution, time, temperature
- Combination of bioassays to characterize the given effect(s)
 - Which assays to use?
- Interpretation and selection (fitness for purpose) of bioassays: Route of exposure, bioavailability of the active compounds, threshold for the effect (for risk assessment/sample prep)
- Quality control (extraction efficiency, normalization of the effect using reference compounds)
- Determination of LOD for a reference compound – to compare sensitivity
- Confirmation procedures (chemical, other bioassays, in-silico etc.)