



Good Food, Good Life

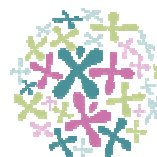
Quantification of milk traces in food matrices using LC/MS

AOAC/ASFILAB workshop, Paris

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Nestlé Research™



Provide safe food with informative labels for food allergic consumers, and minimize precautionary labeling

ELISA technique

- is currently accepted by the authorities and used in Nestlé labs
- selective and sensitive (ppm)
- cross-reactivity possible
- target molecules are not always known
- is fast and easy to use
- little investment costs
- Depended on antibody or kit manufacturers
- is sensitive to protein processing e.g. denaturation and modification
- single-analyte method
- low dynamic range (e.g. 2-20 mg/kg)

LC-MS/MS technique

- is currently evaluated in several labs
- highly selective and sensitive (ppm)
- very specific
- target molecules are known
- is fast, requires MS knowledge
- expensive equipment
- no antibodies required
- not sensitive to protein denaturation, but modification
- Single or multi-analyte formats
- high linear dynamic range (e.g. 2-1000 mg/kg)

Why alternative approaches to ELISA?

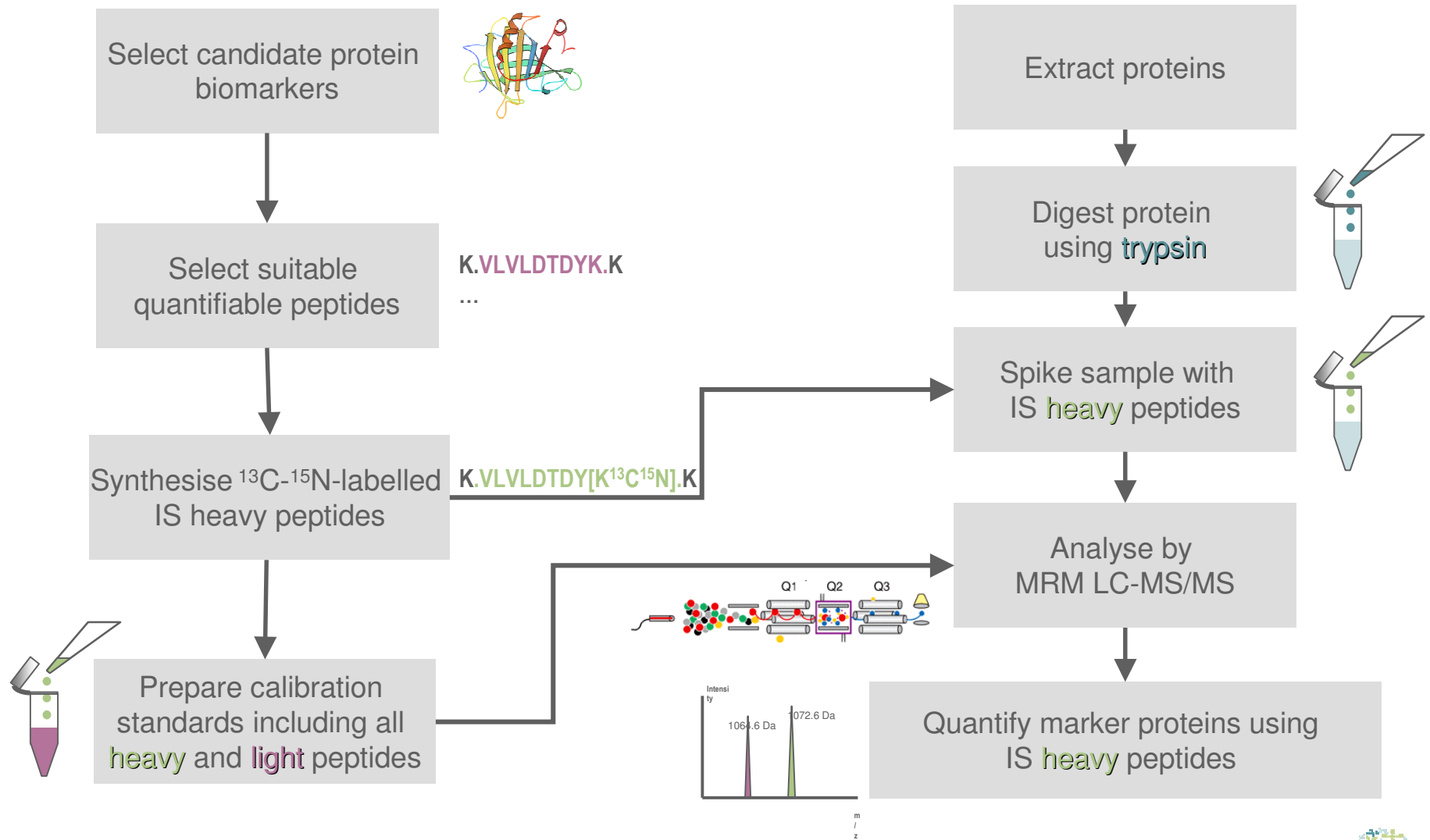
- Need of confirmatory methods
- S/MRM-MS is widely used as the quantification tool of choice for small molecules
- MRM-MS serves as a tool for absolute protein and peptide quantification
- MRM-MS allows detection of multiple unrelated food allergens in a single analysis
- MRM-MS is highly specific
 - allows detection of
 - processed (modified) peptides/proteins
 - peptides/proteins denatured during sample preparation
- Multiplex detection offers control over several peptides/proteins
- Fully transparent
- Offers more flexibility and cost reduction

- Analytical strategy
- Selection of target proteins and peptides
- Application in different matrices
 - Sample preparation
 - Determination of milk traces in food
 - Comparison with ELISA/FAPAS
- Conclusion

Internal standard by isotope dilution

- **Stable isotope-labelled synthetic peptides**
 - Known from peptidomics and phosphorylation studies
- **QconCAT synthetic proteins**
 - Complex mixtures, protein complexes
- **Full-length isotope-labeled proteins (PSAQ)**
 - Accurate quantification on prefractionated samples

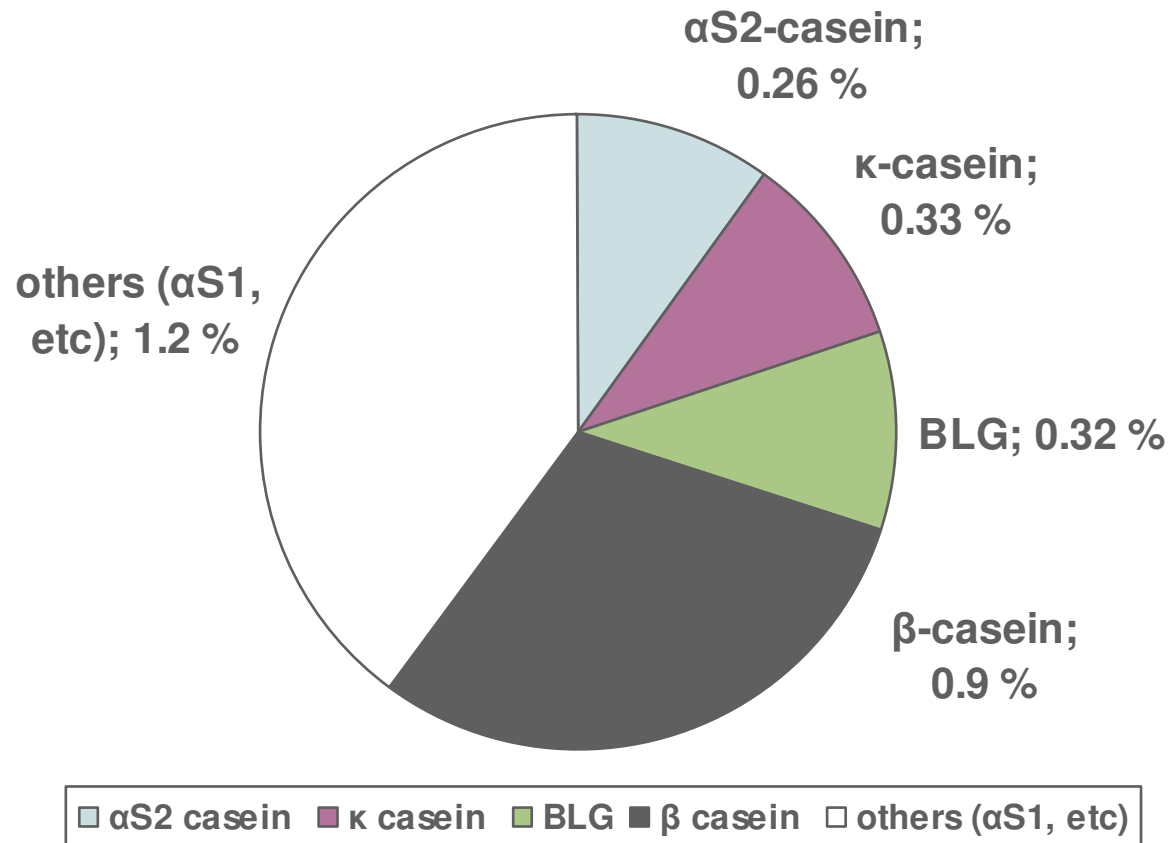
Peptide quantification by isotope dilution – variant I



- Selection of reference protein
 - Specific for milk proteins from different species (cow, buffalo) but not specific for other food ingredients (e.g. egg)
 - Ideally 2-3 reference proteins per allergenic compound
 - Good extraction properties and solubility (no membrane proteins)
 - None to few posttranslational modifications, modifications during food processing
- Selection of marker peptides
 - Sequence specificity
 - No Met, Cys, Trp residues
 - Region with low risk of miscleavages
 - Retention time
 - Ionization capacity
 - 7-15 amino acids/peptide



Distribution of caseins and BLG in total milk protein



The Quantifiers - selection of peptides

- Naturally occurring variants and modifications have to be considered
 - All known variants could be excluded when selecting the quantifier peptides for e.g. beta lactoglobulin (source e.g. SWISS PROT database)

Sequence annotation (Features)						
	Feature key	Position(s)	Length	Description	Graphical view	Feature identifier
Molecule processing						
<input type="checkbox"/>	Signal peptide	1 – 16	16	Ref.6 Ref.8		
<input type="checkbox"/>	Chain	17 – 178	162	Beta-lactoglobulin		PRO_0000017903
Amino acid modifications						
<input checked="" type="checkbox"/>	Disulfide bond	82 ↔ 176		Ref.13 Ref.19		
<input checked="" type="checkbox"/>	Disulfide bond	122 ↔ 137		Alternate Ref.13		
<input checked="" type="checkbox"/>	Disulfide bond	122 ↔ 135		Ref.13 Ref.19		
Natural variations						
<input checked="" type="checkbox"/>	Natural variant	61	1	E → Q in variant D. Ref.10		
<input checked="" type="checkbox"/>	Natural variant	72	1	I → L in variant W. Ref.8		
<input checked="" type="checkbox"/>	Natural variant	75	1	Q → H in variant C; found only in the Jersey breed. Ref.14		
<input checked="" type="checkbox"/>	Natural variant	80	1	G → D in variant A. Ref.6 Ref.1 Ref.9		
<input checked="" type="checkbox"/>	Natural variant	134	1	A → V in variant A. Ref.6 Ref.1 Ref.9 Ref.12		
Experimental info						
<input checked="" type="checkbox"/>	Sequence conflict	121	1	F → V in CAA32835. Ref.1		
<input checked="" type="checkbox"/>	Sequence conflict	136	1	Q → E Ref.12		
Secondary structure						
		1				178
					<input checked="" type="checkbox"/> Helix <input checked="" type="checkbox"/> Strand <input type="checkbox"/> Turn	

Sequence homology

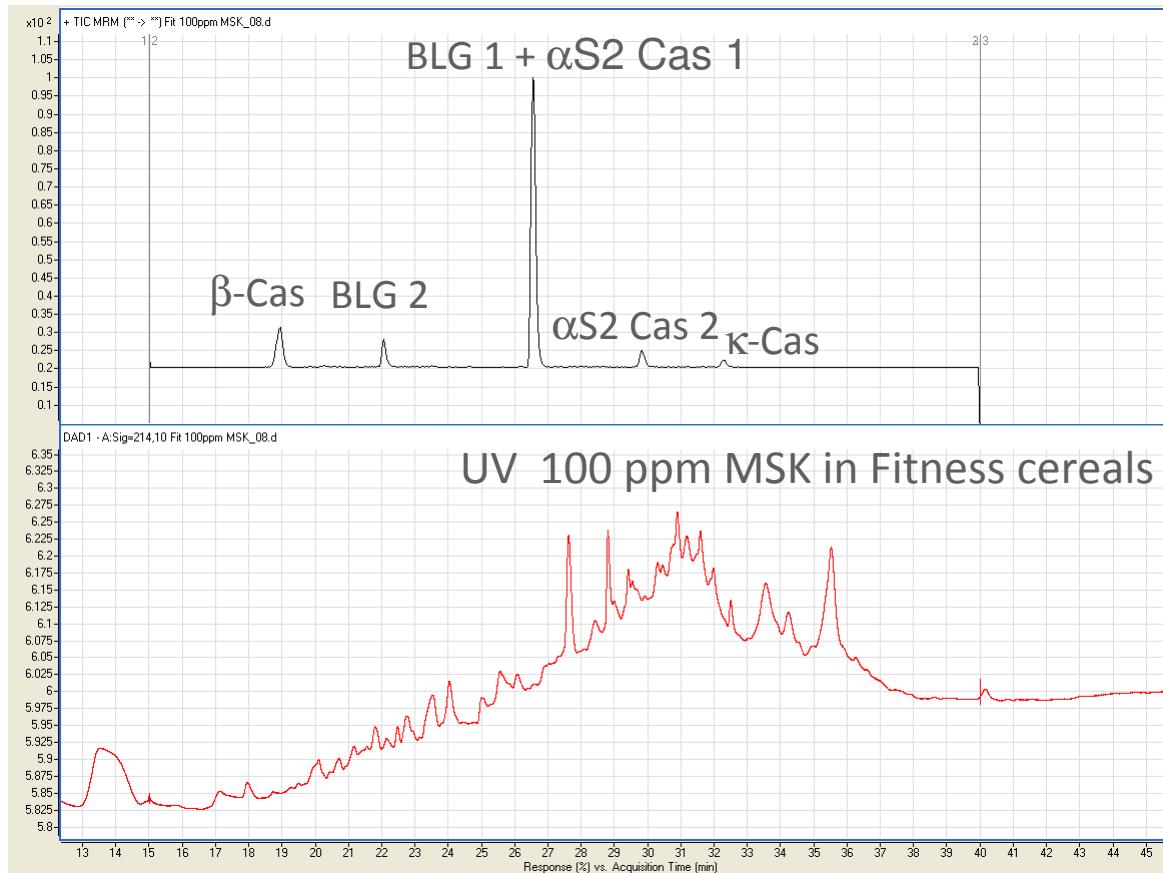
Peptide sequence	Protein	Bovine	Water buffalo	Goat	Sheep	Reindeer	other even-toed ungulate	Human	Bacteria
YIPIQYVLSR	κ -casein	✓	✓	✓	✓	✓	✓	✗	✗
FALPQYLK	α -S2 casein	✓	✗	✗	✗	✗	✗	✗	!!!
ALNEINQFYQK	α S2-casein	✓	✓	✓	✓	✓	n.a.	✗	✗
AVPYPQR	β -casein	✓	✓	✗	✗	✗	✗	✗	1?
VLVLDTDYK	BLG	✓	✓	✓	✓	✓	n.a.	✗	✗
TPEVDDEALEK	BLG	✓	✓	✗	✗	✗	✗	✗	✗

- 5/6 peptides cover both, cow's and buffalo's milk → Asian market !

Typical TIC of selected peptides from milk proteins

- Total ion chromatogram intensities of selected tryptic peptides in MSK spiked Nestlé Fitness cereals

TIC 100 ppm MSK in Fitness cereals



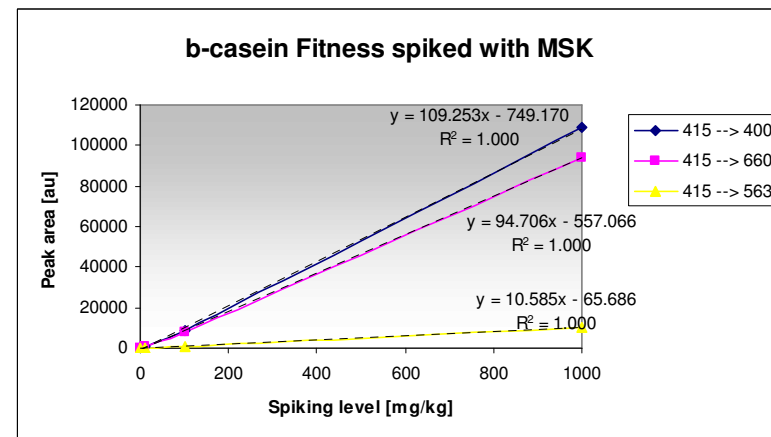
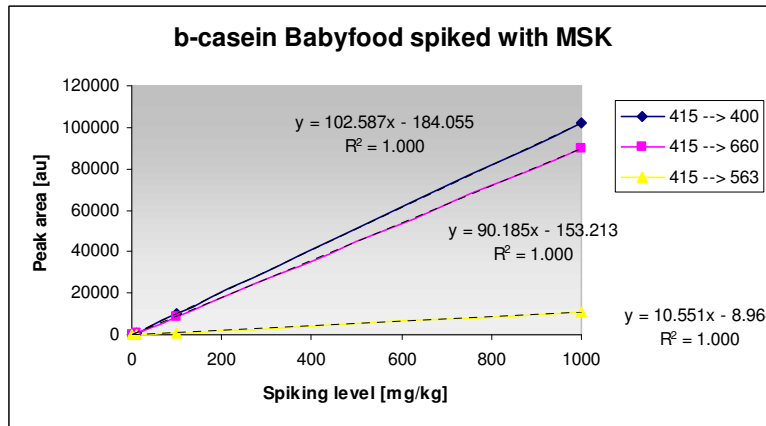
Selection of transitions - CASEIN

- Linearity and intensity of transitions was analysed in blank baby food and cereals and spiked with MSK at 10, 100, and 1000 mg/kg

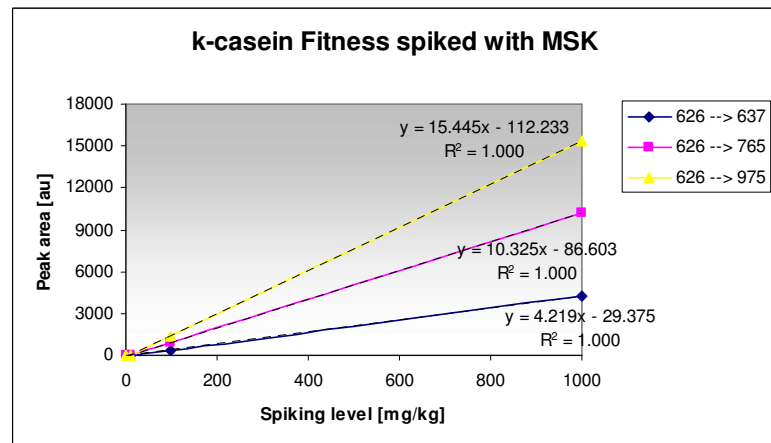
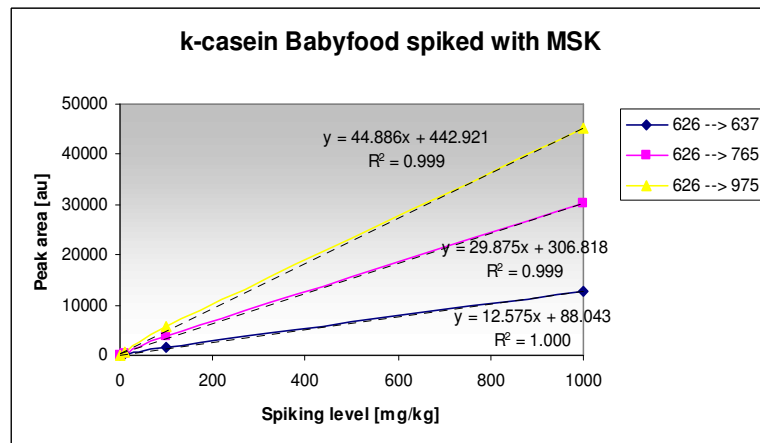
Baby food

Fitness Cereals

β-casein



κ-casein

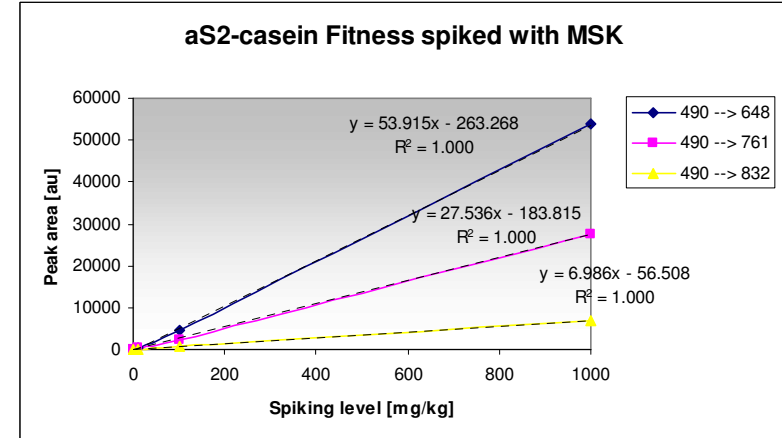
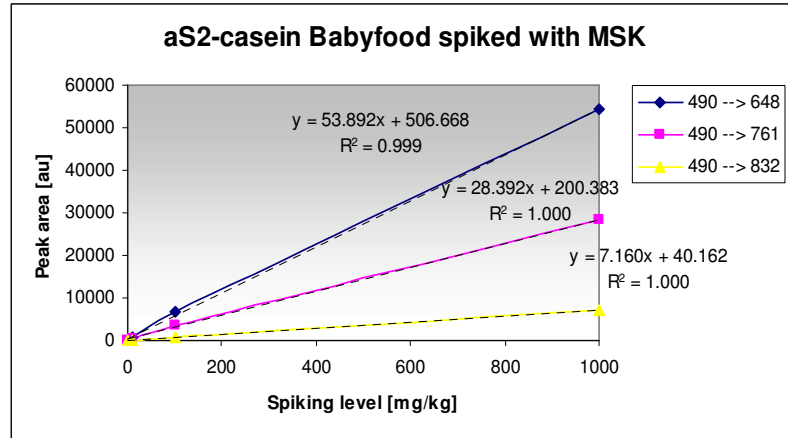


Linearity for different transitions - CASEIN

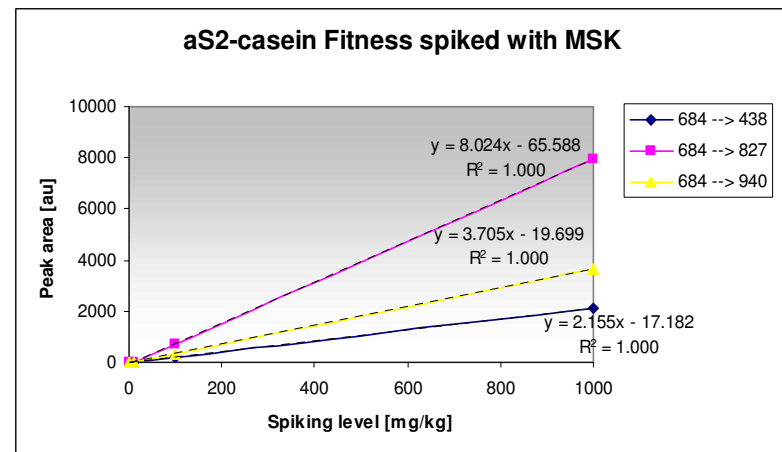
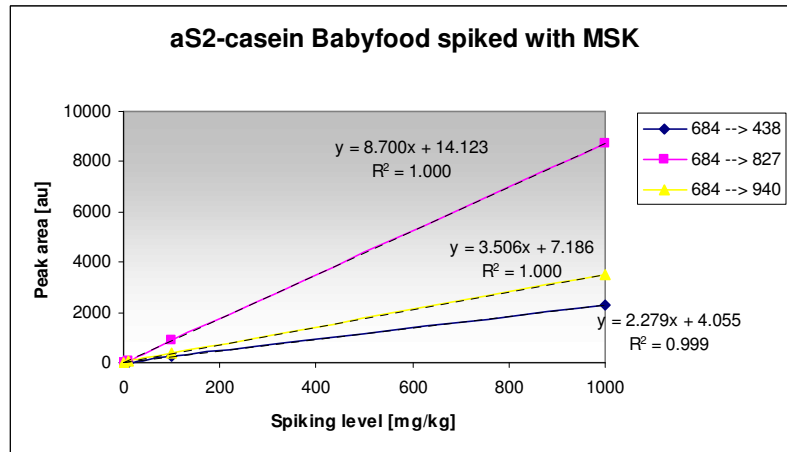
Baby food

Fitness Cereals

αS2-casein

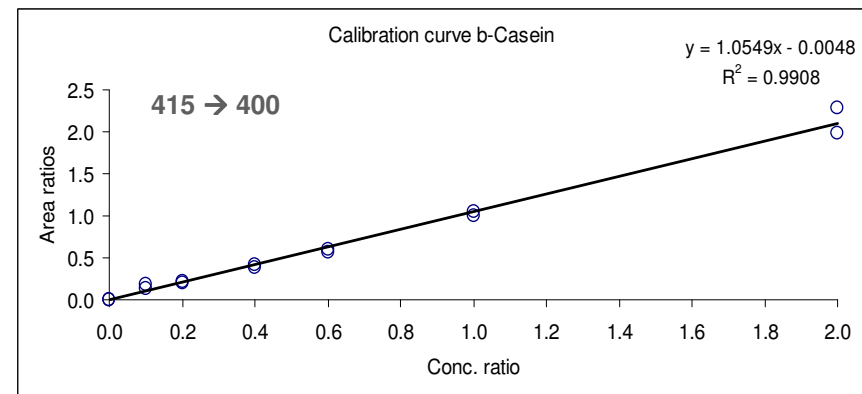
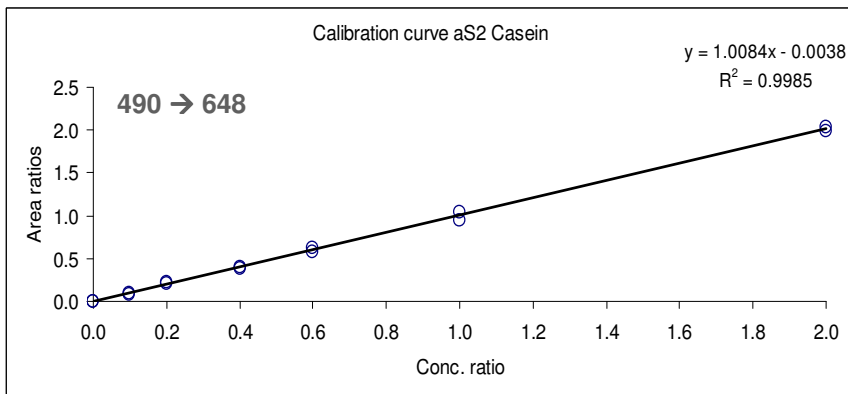
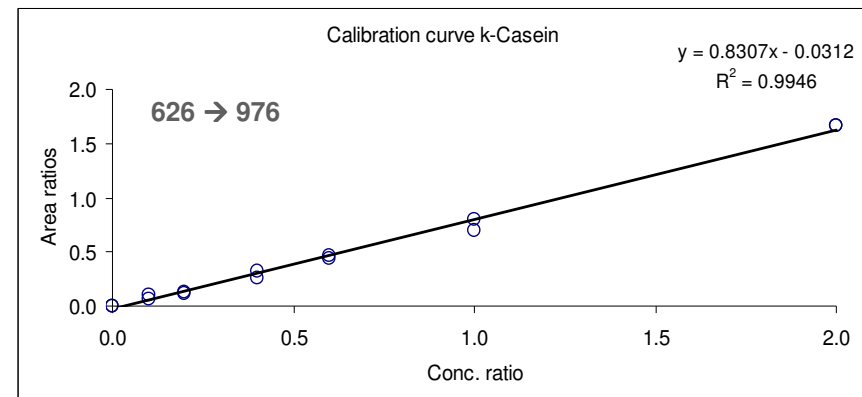
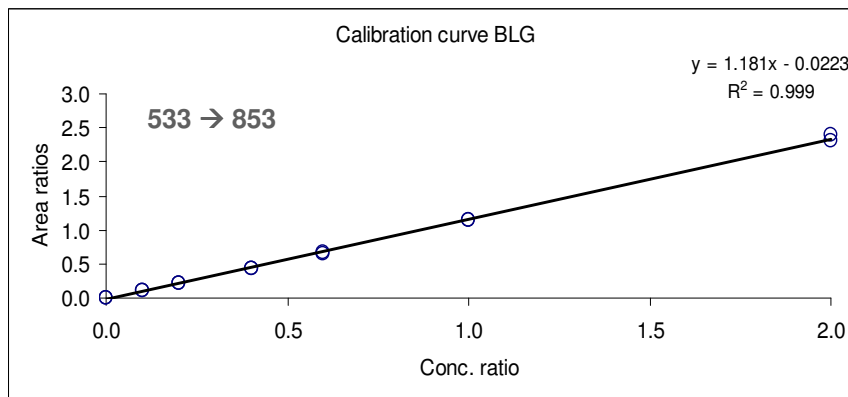


αS2-casein



Calibration Mix contains all IS peptides

- The Calibration Mix contains all IS peptides (**heavy** and **light**) of the four milk proteins
- IS_{heavy} const. 50 fmol; pure standards_{light} from 1-100 fmol (~30-3000 ng/mL MSK)
- Calibration curves follow linear relationship



- ↓ Extract 1g of sample with 10 ml 50 mM NH_4HCO_3 + 1 M urea (60°C).
- ↓ Shake for 15 min at RT, and centrifuge 10 min (5'000 x g and 4°C).
- ↓ Digest an aliquot of supernatant using trypsin at 37°C overnight.
- ↓ If needed, dilute with 0.1% Formic Acid.
- ↓ Mix with Internal Standard (a mixture of 6 labelled peptides).
- ↓ Filter if needed.
- ↓ Injection of 20 μl , corresponds to 50 fmol of each IS (41 to 68 pg, according to the MW of peptide).

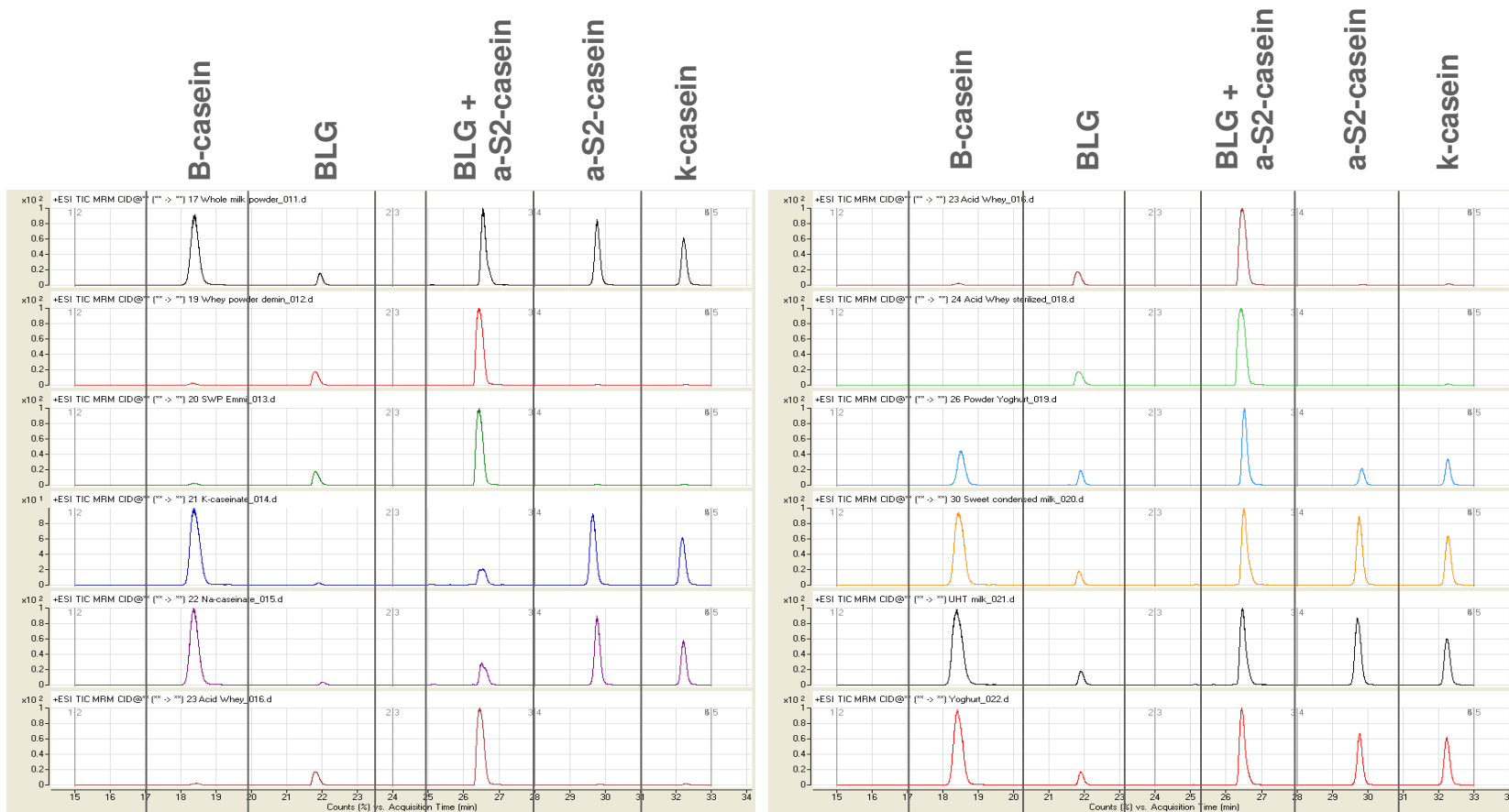
Instrumentation:

1. Agilent HPLC 1200 series coupled to Agilent 6460 mass spectrometer
2. Agilent HPLC 1100 series coupled to Applied Biosystems 4000 Q TRAP® mass spectrometer

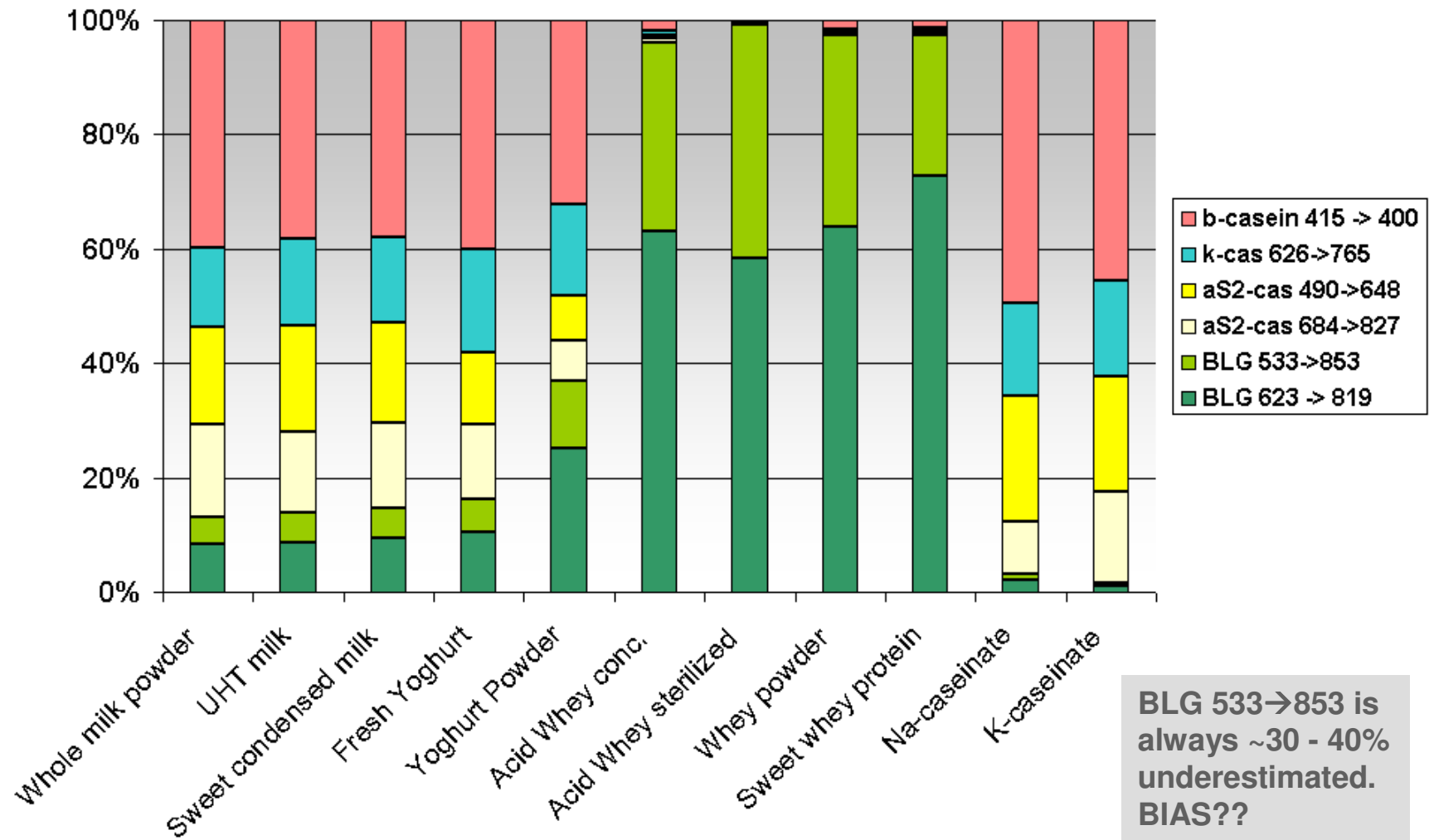
On both, Waters Symmetry300™ C18 3.5 μm , 2.1 x 150 mm, 1 hour gradient,
0.2 ml/min

Effect of processing on detectability of target peptides in typical dairy ingredients

- Selected processed dairy ingredients MRM TICs
 - spray dried, UHT, concentrated, precipitated, defatted, fermented



Ratios of detected milk protein dairy ingredients



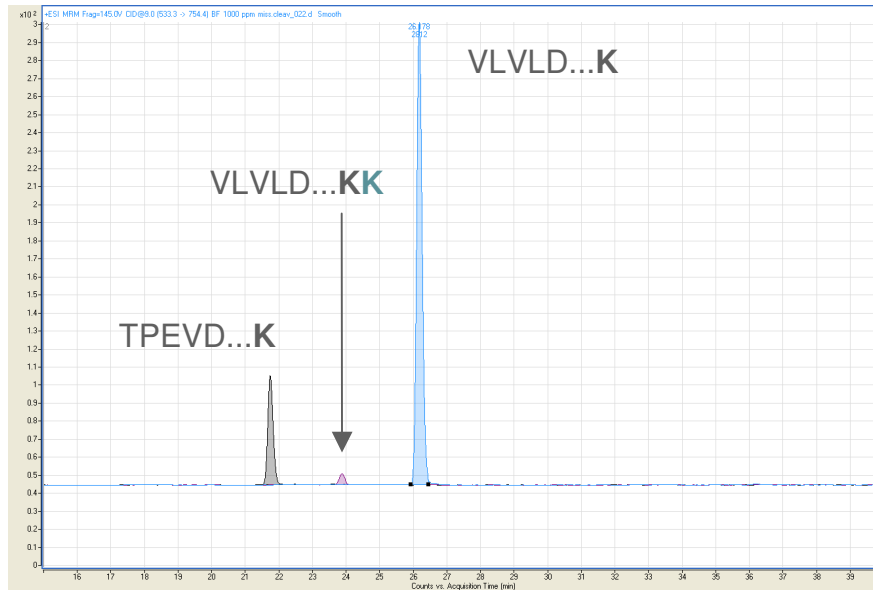
- Matrix components may induce interferences
 - Digestion efficiency
 - pH
 - other proteins
 - protease inhibitors
 - salts, ...
 - modifications: polyphenols, blocked lysins, glycation of amino acids
 - Extraction efficiency
 - lipids
 - protein-protein interaction
 - Peptide mass
 - modifications: oxidation, polyphenols, blocked lysins, glycation of amino acids, phosphorylation, glycosylation
 - digestion efficiency (missed cleavages)
 - adducts (salt ions)



Digestion efficiency in spiked baby food and cereals

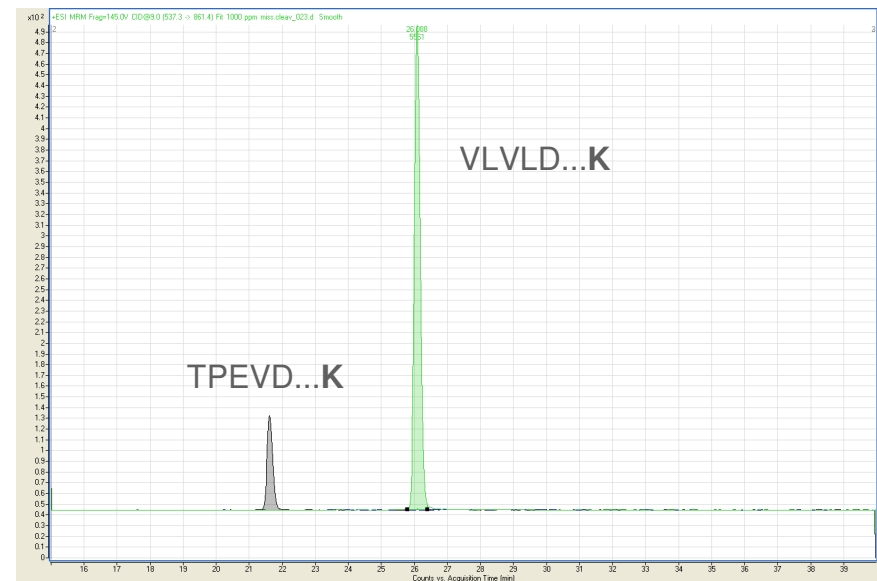
Baby food (1000 ppm MSK)

- Loss dues to **VLVLDTDYKK** is negligible (~ 1.5 area%)
- **TPEVDDEALEKFDK** not present



Cereals (1000 ppm MSK)

- Miscleaved peptides **TPEVDDEALEKFDK** and **VLVLDTDYKK** are not detectable in spiked cereals matrix.



Determination of milk in baby food

Determination of MSK spiked into baby food sample shows good correlation between the three selected casein proteins.

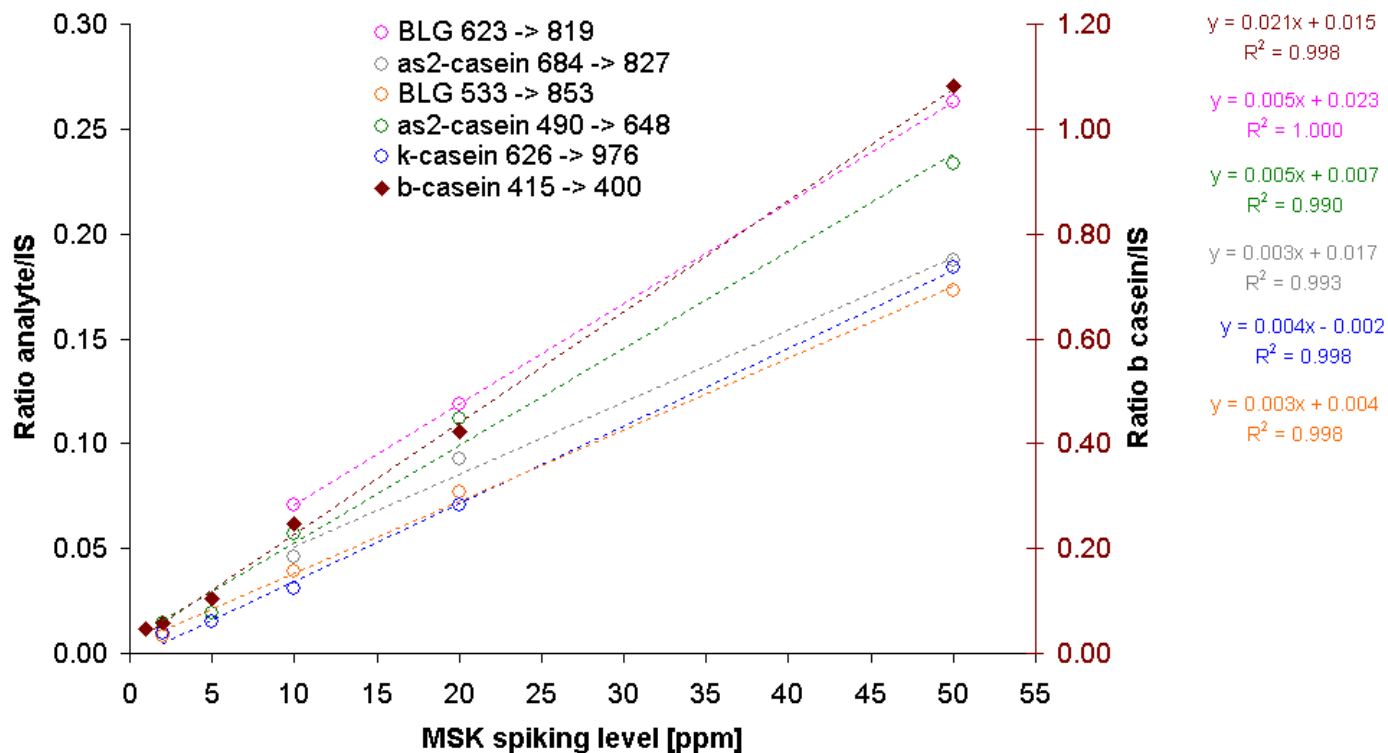
MSK spiking level in baby food	α S2-Casein		κ -Casein		β -Casein	
	MSK μ g/g	% recovery	MSK μ g/g	% recovery	MSK μ g/g	% recovery
0 ppm	n.d.	---	n.d.	---	n.d.	---
10 ppm	7.5	75	n.d.	---	13.9	139
100 ppm filtered	94	94	n.a.	---	79	79
100 ppm	96	96	78	78	78	78
1000 ppm	793	79	824	82	859	86

- Filtration of samples does not effect recovery rates (BF and Fit)
- Filtration of standard solutions reduces amounts to ~60 %

Linearity and LOD/LOQ in infant cereals

Sinlac (milk-free infant cereal, 15.5% total protein) was spiked with 1-50 ppm MSK

- 10 ppm can be quantified based on all reference peptides (based on S/N ratio)
- 2 ppm can be quantified based on β -casein and one BLG peptide (based on S/N ratio)



LOD/LOQ for MSK in infant cereals

	BLG 533 → 853		BLG 623 → 819		α2-casein 684 → 827		α2-casein 490 → 648		β-casein 415 → 400		κ-casein 626 → 976	
	MSK µg/g	Rec. [%]	MSK µg/g	Rec. [%]	MSK µg/g	Rec. [%]	MSK µg/g	Rec. [%]	MSK µg/g	Rec. [%]	MSK µg/g	Rec. [%]
Sinlac blank	nd	--	nd	--	nd	--	nd	--	nd	--	nd	--
1 ppm MSK	nq	--	nd	--	nd	--	nq	--	2.4	240	nq	--
2 ppm MSK	3.1	157	nq	--	nq	--	nq	--	2.8	140	nq	--
5 ppm MSK	4.2	84	3.4	67	nq	--	3.3	66	4.9	98	6.1	122
10 ppm MSK	6.3	63	5.9	59	2.8	28	8.7	87	11.5	115	8.1	81
20 ppm MSK	10.1	51	11.7	59	9.6	48	16.6	83	19.4	97	13.4	67
50 ppm MSK	19.9	40	29.1	58	23.5	47	34.0	68	49.4	99	28.1	56

- LOQ in Sinlac (milk-free infant cereal, 15.5% total protein) is **5 ppm MSK**
- LOD in Sinlac ~ **2 ppm MSK**
- Best recovery rates were obtained for β-casein

Comparison with ELISA – Recovery rates

ELISA

RIDASCREEN Fast BLG

Fitness Cereals	Fit 0 ppm	Fit 10 ppm	Fit 100 ppm	Fit 1000 ppm
recovery	nd	63 %	35 %	34 %

LC-MS/MS

Best transitions

Fitness Cereals	Fit 0 ppm	Fit 10 ppm	Fit 100 ppm	Fit 1000 ppm
recovery BLG 533 → 853	nd	51 %	54 %	56 %
BLG 623 → 819	nd	62 %	66 %	68 %

FAPAS – Beta lactoglobulin

FAPAS 2765	Participants N	BLG in mg/kg Median
ELISA systems	6/9	35.0
R-Biopharm	10/15	148
LC-MS/MS		128 _(533→853) + 189 _(623 → 819)
FAPAS 2751	Participants N	BLG in mg/kg Median
ELISA systems	4/7	18.7
R-Biopharm	17/20	47.6
Tepnel	5/8	20.5
LC-MS/MS		49 _(533→853) + 71 _(623 → 819)

A bias (~0.7) between BLG 533→853 and 623 → 819 was observed analysed ingredients and in both FAPAS samples.

If available, cross contaminating ingredients should be analysed for relative ratios of target peptides.

- 6 tryptic peptides covering BLG and 3 casein proteins were selected
- Linear calibration curves were obtained for 1-100 fmol (~30-3000 ng/mL MSK)
- LC-MS/MS allows determination of processed milk e.g. spray dried, UHT, concentrated, precipitated, defatted, and fermented
- “Simple” sample preparation w/o any enrichment is sufficient
- Method was evaluated in matrices low, medium and high in protein amount
- LOQ in e.g. protein-rich infant cereal is **5 ppm skim milk powder, LOD ~ 2 ppm MSK**
 - This corresponds to LOQ of ~ **0.2 ppm for BLG and respective casein proteins**
 - LOQ is comparable with ELISA
 - Comparison with ELISA results show similar or higher values (dep. on ELISA kit)
 - Recovery rates were found similar or higher compared to ELISA
- Allows direct measurement of defined target molecules

The Team



*Hans
Weymuth*



*Veronique
Parisod*



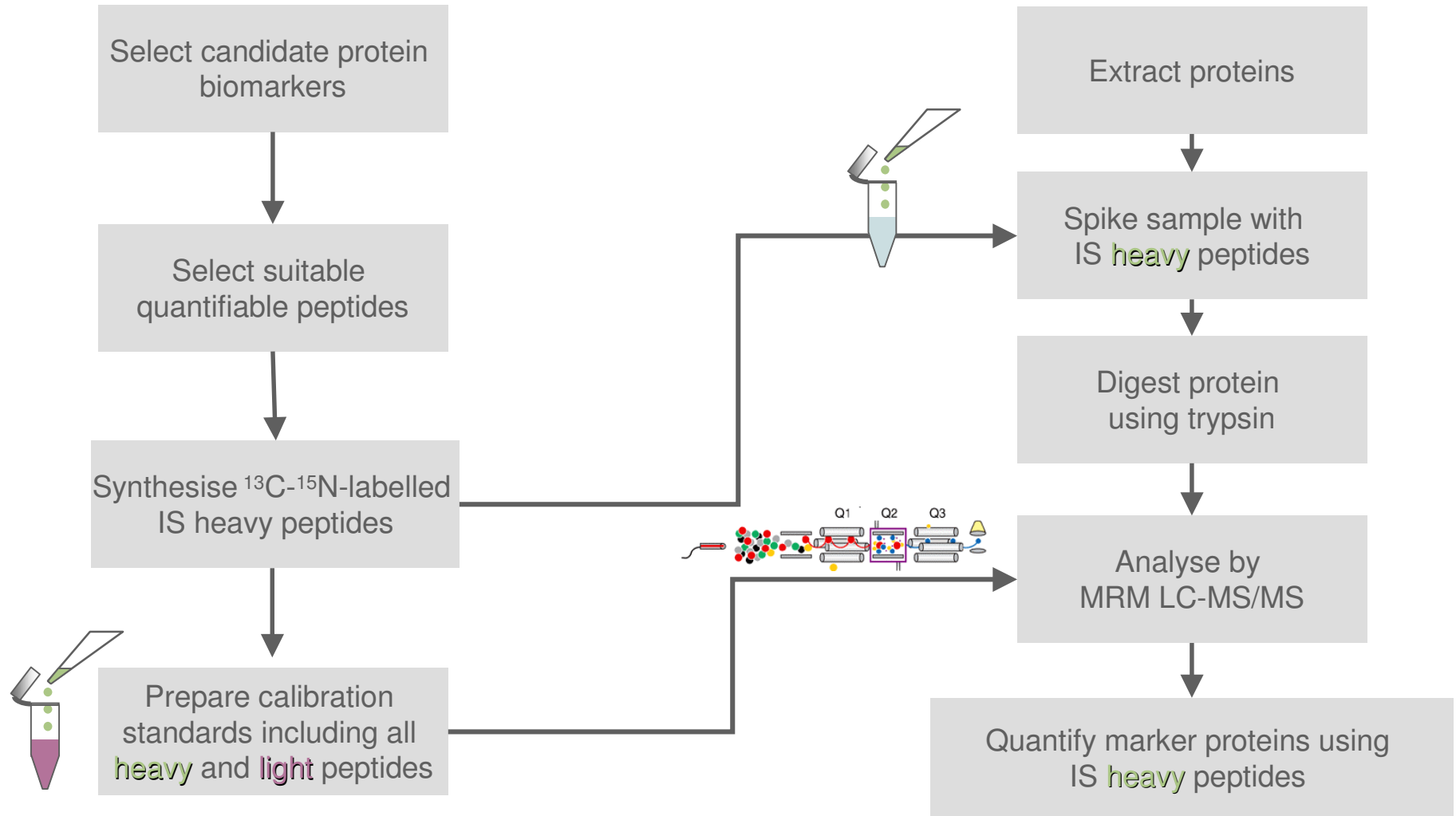
*Petra
Lutter*

Thanks to

*Sylvie Benet (NRC), Mathieu Dubois (NRC), Eric Gremaud (NRC),
Frank Vanrobaeys (Nestlé R&D Beijing)*

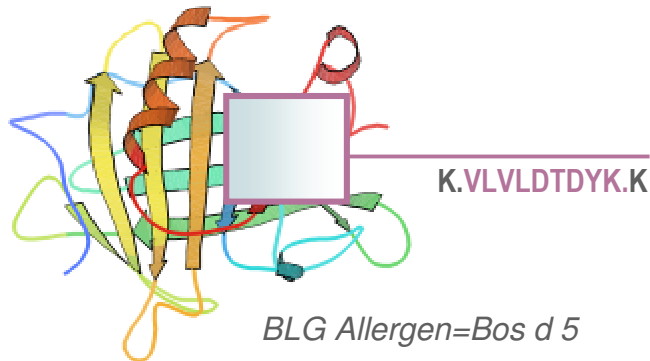


Peptide quantification by isotope dilution – variant II



Absolute protein quantification strategy

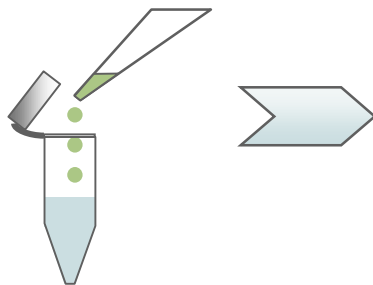
Select allergenic protein



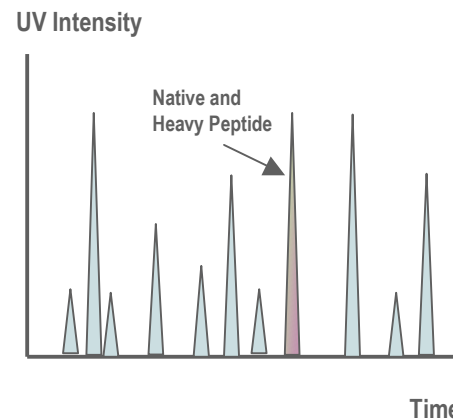
Synthesise heavy peptide analog (IS)



Spike IS into analyte



Separate by HPLC



Quantify by using MS as detector

